

Comprehensive ripeness-index for prediction of ripening level in mangoes by multivariate modelling of ripening behaviour

Vijayaram Eyarkai Nambi^{1*}, Kuladaisamy Thangavel², Annamalai Manickavasagan³, and Sultan Shahir⁴

¹Central Institute of Post-harvest Engineering and Technology, Ludhiana, India

²Horticultural College and Research Institute, Tamil Nadu Agricultural University, Coimbatore, India

³Department of Soils, Water and Agricultural Engineering, Sultan Qaboos University, Sultanate of Oman

⁴T.K.M.Institute of Technology, Kollam, India

Received June 28, 2016; accepted December 8, 2016

Abstract. Prediction of ripeness level in climacteric fruits is essential for post-harvest handling. An index capable of predicting ripening level with minimum inputs would be highly beneficial to the handlers, processors and researchers in fruit industry. A study was conducted with Indian mango cultivars to develop a ripeness index and associated model. Changes in physicochemical, colour and textural properties were measured throughout the ripening period and the period was classified into five stages (unripe, early ripe, partially ripe, ripe and over ripe). Multivariate regression techniques like partial least square regression, principal component regression and multi linear regression were compared and evaluated for its prediction. Multi linear regression model with 12 parameters was found more suitable in ripening prediction. Scientific variable reduction method was adopted to simplify the developed model. Better prediction was achieved with either 2 or 3 variables (total soluble solids, colour and acidity). Cross validation was done to increase the robustness and it was found that proposed ripening index was more effective in prediction of ripening stages. Three-variable model would be suitable for commercial applications where reasonable accuracies are sufficient. However, 12-variable model can be used to obtain more precise results in research and development applications.

Keywords: ripening level, alphonso, banganapalli mango ripening, multi linear regression, partial least square regression, and modelling of ripening

INTRODUCTION

Among climacteric fruits, mango (*Mangifera indica* L.) is one of the most important species. India ranks first in mango production in the world, accounting for 41.5% (FAOSTAT), and the annual production is estimated to be nearly 18 million t (Saxena and Gandhi, 2015). The com-

mercial pack houses and pulping industries need a rapid quality inspection method for the prediction of fruit ripeness level or stage.

Fruit ripening is a highly coordinated, genetically programmed, and irreversible phenomenon involving a series of physiological, biochemical, and organoleptic changes that lead to the development of a soft and edible ripe fruit with desirable quality attributes (Prasanna *et al.*, 2007). The degree of ripeness or level of ripening of climacteric fruits is an important criterion for determining the optimal post-harvest strategies for handling and marketing of the fruits. Changes in biochemical, physiological composition, colour, textural and rheological properties associated with ripening have been reviewed and reported by many researchers (Bashir and Abu-Goukh, 2003; Brady, 1987; Charles and Tung, 1973; Chen and Ramaswamy, 2002; Lizada, 1993; Nambi *et al.*, 2016a, 2016b; Seymour *et al.*, 1993; Stover and Simmonds, 1987).

Padda *et al.* (2011) compared firmness, total soluble solids (TSS) and dry matter to assess the ripening in mangoes using principal component analysis (PCA) and reported that firmness test with penetrometer was suitable to assess changes. Kienzle *et al.* (2012) studied the effect of harvest time on post-harvest quality using multivariate methods. But these studies did not propose any consolidated model or index to quantify or predict ripeness level. Saranwong *et al.* (2004) and Rungpichayapichet *et al.* (2016) reported on the prediction of ripeness and eating quality of mangoes using NIR spectroscopy. All these methods cannot be used

*Corresponding author e-mail: eyarkainambi@gmail.com

as a ready reckoner with minimum inputs, and they need high cost instruments like NIR spectrometer and complex algorithms.

Vásquez-Caicedo *et al.* (2005) proposed a ripening index (RPI) for mango ripeness, combining the fruit firmness with sugar acid ratio. A vast range of RPI values were reported in many studies (Blanes *et al.*, 2015; Kienzle *et al.*, 2012; Rungpichayapichet *et al.*, 2016). Even negative RPI values were found for ripe fruits in our preliminary study (Nambi *et al.*, 2015). So RPI may not be correlated directly with ripening stages, since the vast range of RPI values, including negative numbers, leads to confusion, moreover demarcation of RPI could not be done for different ripening levels.

Ripening class index (Rci) was proposed by Joas *et al.* (2009) based on respiration rate. The Rci does not account for changes of internal and external quality parameters. Jha *et al.* (2007) reported a model for predicting harvest maturity using colour coordinates with the help of maturity index ($I_m = \text{TSS}/8 \cdot 100$) which was developed to assess the appropriate harvest time. The proposed models would not be suitable for ripening, since the model did not deal with the changes during ripening.

Modelling of ripening behaviour would fulfil the need for rapid prediction of ripening level of fruits. Mathematical modelling is gaining interest in predicting chemical, physical and microbiological changes during food processing and storage. It can provide easy prediction and access for further simulation of any process. An index which is capable of explaining the ripening level / behaviour / stage explicitly, with inclusion of all changes occurring in physicochemical, colour and textural properties during fruit ripening, would be more appropriate for ripening prediction. For consolidating the changes in physicochemical, colour and textural properties cumulatively, a higher end statistical approach needs to be adopted while developing such an index. At the same time, the index should be easy to compute and calculate with readily measurable parameters.

Therefore, the objective of this study was to evaluate three multivariate regression techniques (partial least square regression (PLS), principal component regression (PCR) and multi linear regression (MLR)) to predict ripening index (I_R) in three mango varieties using measured quality attributes.

MATERIALS AND METHODS

Three cultivars of mango (Alphonso, Banganapalli and Neelam) were collected at 100-105DFFB (days from full bloom) maturity from two different locations of Tamil Nadu, India, for two seasons (2014 and 2015). Samples collected in 2014 were used in model calibration and those collected in 2015 were used in model validation. The collected mangoes were prepared and kept for ripening as proposed by Nambi *et al.* (2015). From the harvested lot, sound man-

goes were culled out, desapped, washed and shade-dried for 30 min. The selected mangoes were treated with 200 ppm ethylene (0.02%) for 24 h in a ripening chamber at 20°C with 85% RH. After the treatment, the mangoes were kept for ripening at the same temperature and RH (20°C and 85% RH). Three random samples from each variety were taken at 24 h intervals for quality measurement.

The quality parameters, *viz.* TSS, pH, external and internal colour values, titrable acidity (TA) and textural parameters were measured throughout the ripening period. Homogenised mango pulp was obtained with laboratory mixer to determine TSS and TA as reported by Vásquez-Caicedo *et al.* (2005). TSS was measured using a digital refractometer (ATAGO Co Ltd., Japan) in °Brix. The pH value was measured by using a digital pH meter (Systronics µpH system, India). The TA was calculated as grams citric acid equivalent/100 g sample.

The textural characteristics were measured using the Texture Analyzer (TA-HDi, Stable Micro Systems, UK). 4 mm cylindrical probe (P/4) was used with 1 mm s⁻¹ pre-test speed, 0.1 mm s⁻¹ test speed, 1 mm s⁻¹ post-test speed, and 10 mm penetration depth to obtain force displacement curve. Peel strength: PS (maximum force required to penetrate the peel in Newton), stiffness: S (slope of the force/displacement analysis, FDA, curve till peel strength in N mm⁻¹) and flesh firmness: FF (mean force required to penetrate flesh in Newton) were extracted from FDA curve as proposed by Nambi *et al.* (2015) and Camps *et al.* (2005). Colour coordinates were recorded for both external peel colour (L_e^*, a_e^*, b_e^*) and internal pulp colour (L_i^*, a_i^*, b_i^*) of the mango using a HunterLAB colour meter (Hunter Associates Laboratory, Inc. USA) as proposed by Nambi *et al.* (2016b).

As ripening is a complex process, all changes related to physicochemical, colour and textural properties are highly correlated each other, so the index (I_R) could be described as a function of all variables that change during ripening as given in Eq. (1).

$$I_R = f(\text{physicochemical, colour and textural properties}). \quad (1)$$

The ripeness index (I_R) was presumed to be within the range of 0-1 for easy adaptation and interpretation. With the measured quality parameters, hierarchical cluster analysis was used to group the whole data into 5 stages of ripening (Nambi *et al.*, 2015) using JMP (SAS Institute Inc., USA). Among the linkage methods in hierarchical cluster analysis, Ward minimum variance method was employed to maximise the likelihood of variables in each cluster level. Each classified ripening stage was assigned a value as ripeness index (I_R) with 0.2 interval between each stage. MLR, PLS and PCR multivariate analyses with full cross validation were employed to compare predictability of the ripeness index based on goodness of fit parameters.

The function given in Eq. (1) is more complex, since all the quality parameters need to be estimated for the prediction of ripening level. The ultimate objective of this study was to develop I_R with limited variables using simple models without compromising the predictability. Hence the variable reduction procedure was performed based on weightage of each quality parameter in the prediction models. High weightage quality parameters were pooled together and a separate MLR was employed for each group and the predictability of ripening stage were analysed.

The developed ripeness index (I_R) and corresponding regression models were cross validated with the next season fruits (March – June 2015). For each variety, 3 separate sample lots were collected at 7 days intervals from the same location as mentioned above and kept for ripening in order to get different ripening stages. After 18 days of storage (days calculated from the first lot), different lot samples were mixed together and 80 fruits were selected randomly for quality analysis. Besides, 20 fruits were selected randomly from the commercial market at different ripening stages. Commercial market samples were included in the cross validation process in order to validate the model for its prediction efficacy at commercial level. So in total 100 fruits from each variety were selected and all 12 quality parameters were measured.

From the measured quality parameters, I_R values were calculated using identified models and corresponding stage was obtained as predicted ripening stage. The actual ripening stage of each sample was obtained using the properties chart proposed by Nambi *et al.* (2015).

RESULTS AND DISCUSSION

From cluster analysis, the ripening period of each variety was classified into five stages based on the changes occurred in quality parameters using cluster analysis, and the stages were named as unripe (UR), early ripe (ER), partially ripe (PR), ripe (R) and over ripe/decay (OR) as proposed by Nambi *et al.* (2015). The dendrogram obtained from cluster analysis for each variety is given in Fig. 1.

Three multivariate regression analyses (PLS, PCR and MLR) were used for prediction and compared for their effectiveness in prediction with R^2 , RMSE, standard error and bias of calibration and validation. Obtained results from multivariable regression are given in Table 1. All three regression analyses predicted the ripening stage with higher R^2 value, lower RMSE, standard error and bias in both calibration and validation (Table 1). Comparatively, MLR showed the best results among the three analyses for all the cultivars, with lowest RMSE, SE and bias values. Similar results were reported by Jha *et al.* (2007) in mango for harvest maturity prediction using colour values. This may be due to the lower number of variables. PLS and PCR would be suitable for a higher number of variables than the observation. Hence, MLR was selected for further studies. Moreover, multi linear regression would be easy for further prediction than other models. The regression coefficients are given in Table 2.

For making a more simple and effective model from the 12 variable MLR model, the variable reduction process was carried out based on the weightage of each quality

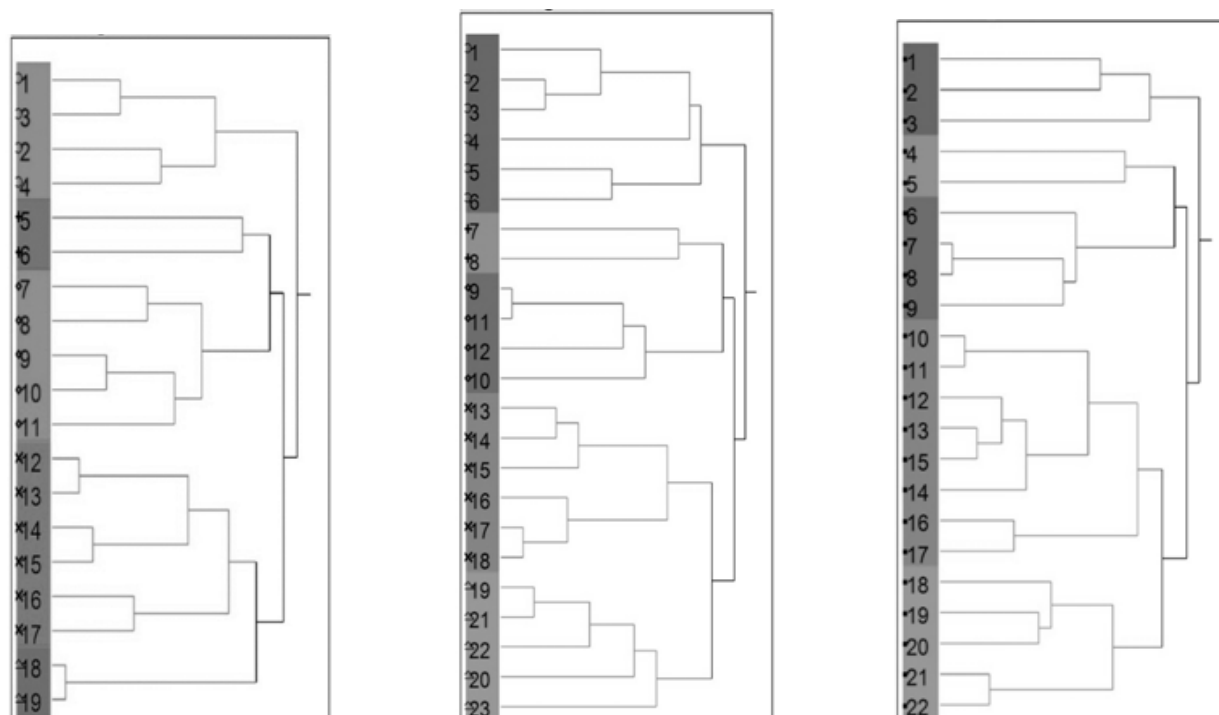


Fig. 1. Dendrograms obtained in Hierarchical cluster with five ripeness levels in Alphonso, Banganapalli and Neelam mangoes (number in the dendrogram indicates the total post-harvest life of respective fruit variety in days).

Table 1. Comparison of various multivariable regression for ripening prediction

Mango variety	Regression type	R ²		RMSE		SE		Bias	
		calibration	validation	calibration	validation	calibration	validation	calibration	validation
Alphonso	PLS	0.985	0.914	0.029	0.050	0.030	0.051	0.000	0.004
	PCR	0.961	0.948	0.051	0.628	0.052	0.064	0.000	-0.003
	MLR	0.990	0.967	0.026	0.076	0.027	0.078	0.000	-0.003
Banganapalli	PLS	0.972	0.958	0.050	0.065	0.051	0.066	0.000	-0.001
	PCR	0.966	0.909	0.055	0.063	0.056	0.064	0.000	0.001
	MLR	0.989	0.960	0.031	0.061	0.031	0.092	0.000	-0.001
Neelam	PLS	0.976	0.971	0.040	0.046	0.041	0.047	0.000	0.001
	PCR	0.974	0.899	0.042	0.048	0.043	0.050	0.000	0.001
	MLR	0.991	0.969	0.025	0.043	0.026	0.085	0.000	0.001

PLS – partial least square regression, PCR – principle component regression, MLR – multi linear regression, R² – coefficient of determination, RMSE – root mean square error, SE – standard error.

Table 2. Regression coefficients of selected models for ripening prediction

Variable	Inter	L* _e	a* _e	b* _e	L* _i	a* _i	b* _i	pH	TSS	TA	PS	S	FF
Alphonso													
All	0.631	-0.001	0.004	0.007	-0.010	-0.001	-0.001	0.068	0.005	-9.041	0.007	-0.001	-0.013
External colour	0.656	-0.016	0.013	0.013									
Textural properties	0.855										-0.021	-0.005	0.026
TSS and acidity	0.139								0.038	-19.615			
Banganapalli													
All	1.288	0.011	-0.001	0.006	-0.019	-0.010	-0.002	0.090	0.005	-33.037	0.001	-0.016	0.001
External colour	-1.451	0.023	0.005	0.012									
Textural properties	0.962										-0.008	-0.017	0.003
TSS and acidity	0.656								0.036	-90.836			
Neelam													
All	1.987	-0.006	-0.001	-0.001	-0.018	-0.002	0.001	0.120	-0.002	-9.888	0.000	-0.003	-0.004
External colour	-0.449	0.015	0.010	0.001									
Textural properties	0.776										-0.007	-0.013	-0.001
TSS and acidity	-0.157								0.044	-15.816			

Inter – intercept, TSS – total soluble solids, TA – titrable acidity, PS – peel strength, S – stiffness, FF – flesh firmness.

Table 3. Weightage (t-values) of each quality parameter in MLR model in ripening prediction

No. variables	Parameters	L* _e	a* _e	b* _e	L* _i	a* _i	b* _i	pH	TSS	TA	PS	S	FF
Alphonso													
12	All	-0.12	0.99	1.46	-1.93	-0.26	-0.30	1.97	0.42	-2.07	1.79	-0.08	-2.16
6	Colour properties	5.59	-0.76	2.18	3.17	-3.75	0.05	-0.50					
3	External colour	-2.72	4.74	3.19									
3	Internal colour				-5.59	4.47	-3.45						
3	Textural properties										-4.34	-0.39	2.32
3	pH, TSS and acidity							3.88	4.50	-5.14			
2	TSS and acidity								10.16	-6.95			
Banganapalli													
12	All	3.47	-0.50	3.35	-3.83	-3.74	-2.63	3.61	1.78	-4.43	0.42	-4.42	0.13
6	Colour properties	5.80	0.00	3.23	-4.79	-2.75	0.36						
3	External colour	4.07	4.55	4.27									
3	Internal colour				-3.12	-2.11	1.31						
3	Textural properties										-5.60	-1.06	0.20
3	pH, TSS and acidity							1.87	3.28	-3.39			
2	TSS and acidity								5.37	-10.08			
Neelam													
12	All	-1.40	-0.40	-0.34	-5.66	-0.34	0.33	2.94	-0.36	-2.33	-0.16	-0.29	-1.56
6	Colour properties	-0.09	3.93	-1.11	-5.11	1.79	0.12						
3	External colour	2.34	3.00	0.35									
3	Internal colour				-5.12	1.66	1.20						
3	Textural properties										-1.64	-0.75	-0.15
3	pH, TSS and acidity							2.07	1.66	-2.43			
2	TSS and acidity								6.07	-3.85			
2	TSS and acidity								2.66	9.62			

Explanations as in Table 2.

parameter on the prediction. Properties like colour coordinates (6 parameters), textural characteristic (3 parameters) and physicochemical properties (TSS, pH and acidity) were grouped as individual groups and evaluated in prediction. The weightage of each quality parameter was found based on t-values is given in Table 3. The goodness of fit

parameters for the 12 variable MLR model are given in Table 4, and higher R² (>0.94) was found with the 12 variables model.

While using MLR with all the colour coordinates (6 parameters) together for prediction, external L* value was found to be a highly influencing parameter, followed by internal L* then by other coordinates in Alphonso and

Table 4. Goodness of fit parameters at different level of variable while using MLR for ripening prediction

Model	No. variables	Quality parameters	Alphonso			Banganapalli			Neelam		
			R ²	RMSE	χ^2	R ²	RMSE	χ^2	R ²	RMSE	χ^2
MLR	12	All	0.962	0.061	0.004	0.992	0.030	0.001	0.936	0.064	0.004
	6	Colour properties	0.947	0.068	0.005	0.969	0.056	0.003	0.919	0.077	0.006
	3	External colour	0.919	0.082	0.007	0.951	0.068	0.005	0.808	0.128	0.016
	3	Internal colour	0.928	0.077	0.006	0.883	0.105	0.011	0.812	0.102	0.010
	3	Textural properties	0.860	0.114	0.013	0.929	0.082	0.007	0.801	0.129	0.017
	3	pH, TSS and acidity	0.937	0.072	0.005	0.896	0.112	0.013	0.828	0.101	0.010
	2	TSS and Acidity	0.929	0.081	0.006	0.889	0.114	0.013	0.813	0.123	0.015
	2	External L* and a*	0.910	0.08	0.006	0.927	0.080	0.007	0.807	0.122	0.015
2nd order	2	External a* and b*	0.916	0.083	0.007	0.934	0.078	0.006	0.796	0.126	0.016
Linear	1	TSS	0.846	0.110	0.012	0.642	0.181	0.033	0.658	0.136	0.019
Quad	1		0.860	0.106	0.085	0.651	0.179	0.027	0.670	0.119	0.010
Linear	1	Acidity	0.764	0.136	0.019	0.797	0.136	0.018	0.561	0.154	0.024
Quad	1		0.770	0.130	0.017	0.801	0.117	0.010	0.560	0.174	0.001
Linear	1	External a	0.704	0.176	0.020	0.770	0.160	0.016	0.541	0.184	0.029
Quad	1		0.710	0.180	0.017	0.780	0.161	0.010	0.540	0.184	0.028
Linear	1	PS	0.808	0.122	0.015	0.915	0.088	0.008	0.698	0.128	0.016
Quad	1		0.810	0.100	0.110	0.919	0.087	0.004	0.710	0.087	0.011

MLR – multi linear regression, Quad – quadratic model, R² – coefficient of determination, RMSE – root mean square error, χ^2 – Chi-square value.

Banganapalli (Table 3). This may be due to a change in external peel colour from dull green to bright yellowish red, and a change in internal colour from bright white to dark yellow during ripening. But a different scenario was observed in Neelam, for which the internal L* value was found as highly influencing a parameter, followed by the external a* value. This might be due to non-uniform colour change that occurred on the peel of Neelam mangoes. From the Table 4, it could be observed that prediction of ripening stage using the 6 colour variables model was more effective, with higher R² (>0.91) and lower RMSE

and chi-squared values in all of the mango cultivars. The regression coefficients of the six-variable model are given in Table 2.

Further to reduce the variables, MLR with 3 variables was evaluated using internal and external colour coordinates as separate groups. Unlike the 6 parameter model, the external a* value had higher influence on prediction than L* and b* in all cultivars of mango while using external colour for prediction. On the other hand, while using internal colour for prediction, L* exhibited a higher influence, followed by a* and b* in all the three mango cultivars.

Multi linear regression models from both internal and external colour groups exhibited similar goodness of fit results in prediction in all mango cultivars (Table 4). From these findings, the 3-variable MLR model with the external colour coordinates would be suitable rather than with the internal colour coordinates, since external colour coordinates can be measured in non-destructive way. The regression coefficients of the 3-variable model (external colour coordinates) are given in Table 2.

Among the three textural characteristics, peel strength was found to be the most influential variable in all the mango cultivars. Flesh firmness was found to be the second influential variable in Alphonso mangoes, which may be due to physiological variations between cultivars which lead to fast softening of fruit pulp in Alphonso compared to Banganapalli and Neelam mangoes. From Table 4, it could be noticed that MLR model based on textural characteristics would be more suitable to predict the ripening stages in Banganapalli, with higher R^2 values (>0.90) and lower RMSE and chi-squared values than the other two mango cultivars. The regression coefficients of the MLR model based on three textural characteristics are given in Table 2.

Among the three physicochemical properties (TSS, pH and acidity), acidity was found to be a highly influential variable, followed by TSS in Alphonso and Neelam. But in Banganapalli mangoes both TSS and acidity exhibited on par influence in the prediction model. Using MLR with three variables (physicochemical properties), effective prediction of ripening stages was achieved in Alphonso and Banganapalli, with higher R^2 value (>0.90), lower RMSE and chi-squared values. In the case of Neelam, prediction was not on par with the other two cultivars ($R^2 < 0.90$). This may be due to physiological variations between the cultivars. The regression coefficients of the MLR model with TSS, pH and acidity are given in Table 2.

For further reduction in variables, two highly influential variables from MLR models containing external colour coordinates (L^* and a^*) and physicochemical properties (TSS and acidity) were selected and evaluated in prediction. The model with TSS and acidity exhibited on par result with the three variable MLR model which contain TSS, acidity and pH. Hence, the two variable MLR model with TSS and acidity would be suitable for prediction of ripening stages in all mango cultivars rather than the three variable MLR model.

The MLR model with external L^* and a^* showed better results in prediction. A second order polynomial model with the external a^* and b^* as proposed by Jha *et al.* (2007) was also evaluated and found to produce on par results with the model containing external L^* and a^* colour coordinates. Though, these two variable models exhibited contending results (Table 4) in the prediction, MLR model containing three external colour coordinates would be more suitable, since the model exhibited more effective results than other

colour based models. Moreover, all the three colour coordinates (L^* , a^* and b^*) can be measured simultaneously in the same device and they can be used together.

In order to make the prediction model most simple, highly influential variables from the three variable models were selected and evaluated for their predictability of ripening stage. Totally four quality parameters (TSS, acidity, external a^* and PS) were selected for single variable evaluation both in linear and quadratic form. Higher order models did not yield fruitful results with a single variable, hence the modelling was limited to 2nd order (quadratic) form. Though acidity was a highly influential variable among TSS, pH and acidity, TSS was also selected for single variable evaluation, since TSS is an easy as well as readily measurable parameter with handheld refractometer. The model containing TSS was found comparatively better in predicting ripening stage in Alphonso mangoes than in the other cultivars. Peel strength based model was found better in ripening prediction in Banganapalli than in the other cultivars. Overall, single variable models did not perform effectively in prediction and yielded lower R^2 values (Table 2). Therefore, single variable models should be avoided for prediction of ripening stages.

Ripening index and RPI were compared for their suitability and effectiveness using observed values from the cross validation set (100 fruits). All 12 quality parameters were measured and I_R values were calculated using the 12 parameter MLR model. RPI was also calculated using the existing formulae. Comparison of RPI with the proposed I_R is given in the form of box plot with outliers (Fig. 2). Similar trend of RPI values was observed in all three cultivars. From Fig. 2a, overlaps could be observed between various ripening stages in RPI values, especially adjacent ripening stages like UR with ER and PR with R. Around 75% of RPI values of UR and ER stages were found overlapping with each other. Likewise, on par RPI values of PR and R stages were found in Banganapalli and Neelam.

Higher variation in RPI values in the last stage (OR) was observed in Alphonso (Fig. 2a). This may be due to a decrease in TSS, especially in Alphonso, observed towards the end of ripening, where decay of fruit starts. From these results it could be concluded that RPI could only be used to predict whether the mango is raw (higher RPI values represents unripe mango) or ripe (lower RPI values represents ripe mango), rather than to predict different stages/ levels of ripening.

On the other hand, the proposed I_R provided clear demarcation between ripening stages without any overlapping between the stages (Fig. 2b). Moreover, using I_R a ripening stage can be easily identified emphatically, since the I_R varies within the defined range of values (0-1).

From the variable reduction studies, best resulting and suitable models were taken for external cross validation with the next season mangoes (March-June, 2015). The

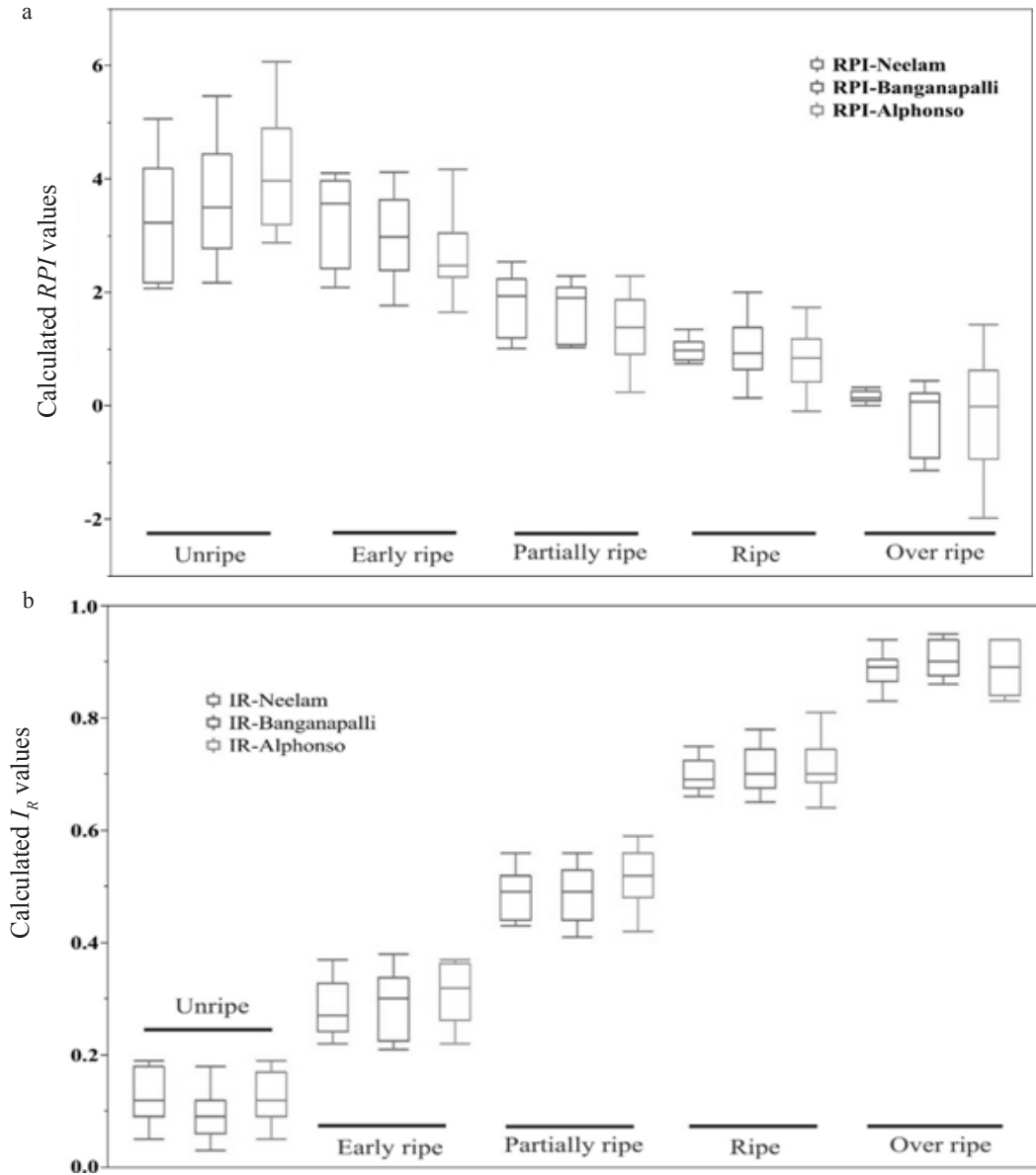


Fig. 2. Comparison of RPI and I_R with five ripening stages for three mango varieties: a – box plot for RPI values, b – box plot for I_R values.

actual and predicted ripening stages/levels were compared and plotted based on the number of correct predictions in each variety (Fig. 3).

While using MLR with all 12 quality parameters, 100% correct prediction was achieved in Alphonso and Banganapalli mangoes, but in Neelam around 99% correct prediction was achieved. In external cross validation, the models with colour coordinates exhibited effective prediction. Either of the 6-variable (all internal and external colour) model or the 3-variable (external colour) model exhibited similar results in cross validation and effectively predicted the ripening stages of all mango cultivars with more than 98% correct prediction (Fig. 3). Hence it could be concluded that instead of the 6-variable complex MLR, the 3-variable model (Eq. (2)) containing external colour

coordinates would be sufficient for predicting ripening stage. Another three variables model with textural characteristics was also tested in the cross validation and the results are shown in Fig. 3. Though the MLR model with 3 textural characteristics resulted in good prediction, the MLR model with 3 external colour coordinates would be more suitable, since colour coordinates are easy to measure with hand held colorimeter and comparatively cost effective compared to texture analysis, moreover the model with external colour exhibited better results than the model with textural parameters.

$$I_R = \text{Intercept} + x L_e^* + y a_e^* + z b_e^*, \quad (2)$$

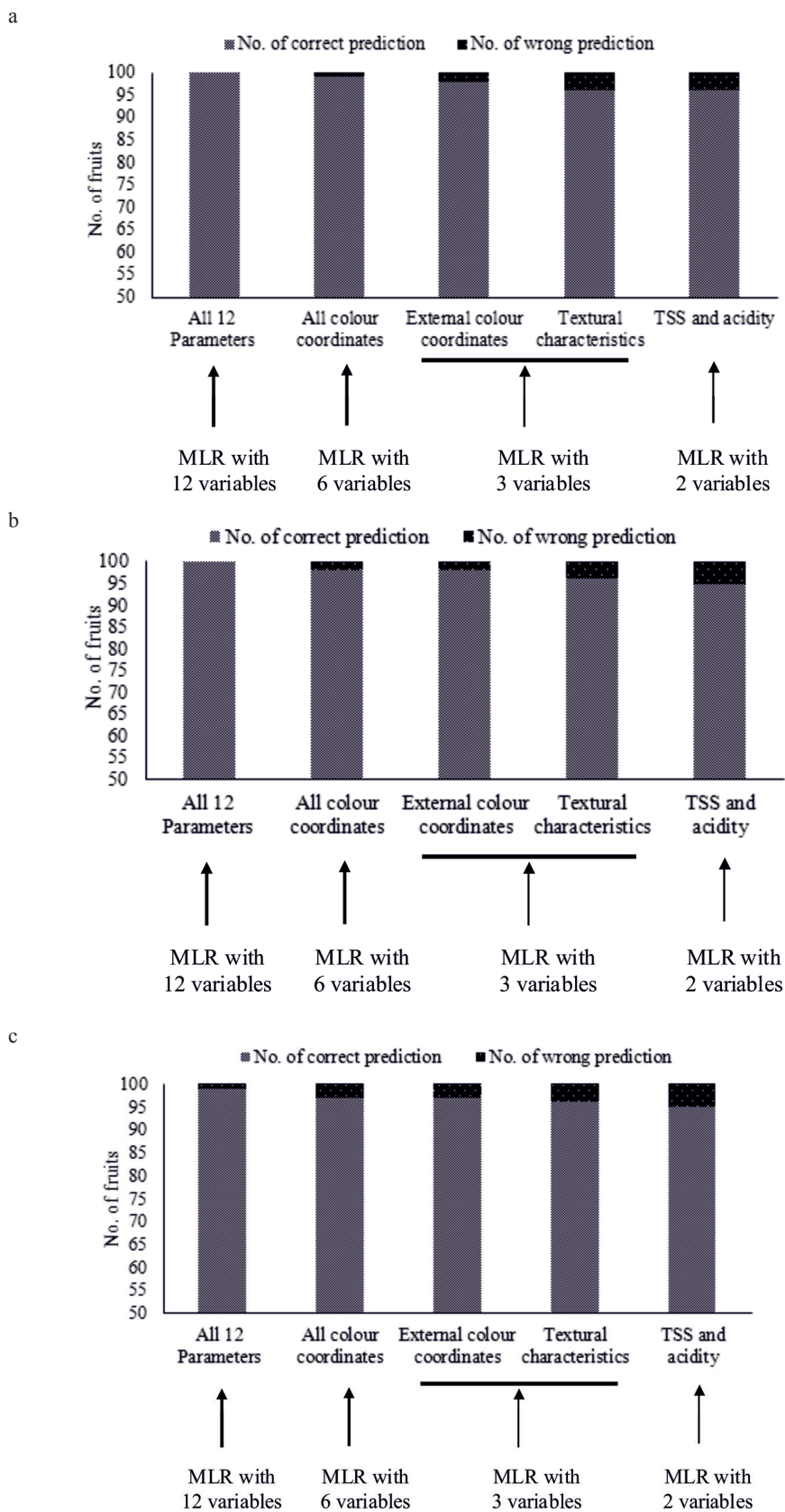


Fig. 3. Results of cross validation with No. of fruit correctly predicted in: a – Alphonso, b – Banganapalli, and c – Neelam mangoes.

$$I_R = \text{Intercept} + x \text{ TSS} + y \text{ TA}, \quad (3)$$

where: x , y and z are corresponding coefficients for the corresponding quality parameters (Table 2).

More than 95% correct prediction was achieved with the two variables model using TSS and acidity. Hence the model (Eq. (3)) with TSS and acid content could be very useful for quick and easy prediction which needs minimum inputs and at the same time produces comparable results to those that can be achieved with the 12 variable MLR model.

CONCLUSIONS

1. The developed index and its multilinear regression models were found effective in the prediction of ripeness level or stage in all the three cultivars of mango.

2. The developed models were further simplified using variable reduction techniques, for rapid and easy prediction.

3. Better prediction was achieved using multi linear models with 2 variables (total soluble solids and acidity) and 3 variables of colour (external L^* , a^* and b^*) coordinates.

4. These simple models would be suitable for processors and other commercial applications where reasonable accuracy is required. However, the accuracy could be improved with 12 quality parameters wherever required.

Conflict of interest: The Authors do not declare conflict of interest.

REFERENCES

- Bashir H.A. and Abu-Goukh A.-B.A., 2003.** Compositional changes during guava fruit ripening. *Food Chemistry*, 80, 557-563.
- Blanes C., Cortés V., Ortiz C., Mellado M., and Talens P., 2015.** Non-destructive assessment of mango firmness and ripeness using a robotic gripper. *Food and Bioprocess Technology* 8, 1914-1924.
- Brady C., 1987.** Fruit ripening. *Annual Review of Plant Physiol.*, 38, 155-178.
- Camps C., Guillermin P., Mauget J., and Bertrand D., 2005.** Data analysis of penetrometric force/displacement curves for the characterization of whole apple fruits. *J. Texture Studies*, 36, 387-401.
- Charles R. and Tung M., 1973.** Physical, rheological and chemical properties of bananas during ripening. *J. Food Sci.*, 38, 456-459.
- Chen C. and Ramaswamy H., 2002.** Color and texture change kinetics in ripening bananas. *LWT-Food Sci. Technol.*, 35, 415-419.
- Jha S., Chopra S., and Kingsly A., 2007.** Modeling of color values for nondestructive evaluation of maturity of mango. *J. Food Eng.*, 78, 22-26.
- Joas J., Caro Y., and Lechaudel M., 2009.** Comparison of post-harvest changes in mango (*cv. Cogshall*) using a Ripening class index (Rci) for different carbon supplies and harvest dates. *Postharvest Biol. Technol.*, 54, 25-31.
- Kienzle S., Sruamsiri P., Carle R., Sirisakulwat S., Spreer W., and Neidhart S., 2012.** Harvest maturity detection for 'Nam Dokmai# 4' mango fruit (*Mangifera indica* L.) in consideration of long supply chains. *Postharvest Biol. Technol.*, 72, 64-75.
- Lizada C., 1993.** Mango. In: *Biochemistry of fruit ripening* (Eds G.B. Seymour, J.E. Taylor, G.A. Tucker), Springer Press, Netherlands.
- Nambi V.E., Thangavel K., and Jesudas D.M., 2015.** Scientific classification of ripening period and development of colour grade chart for Indian mangoes (*Mangifera indica* L.) using multivariate cluster analysis. *Scientia Horticulturae*, 193, 90-98.
- Nambi V.E., Thangavel K., Rajeswari K.A., Manickavasagan A., and Geetha V., 2016a.** Texture and rheological changes of Indian mango cultivars during ripening. *Postharvest Biol. Technol.*, 117, 152-160.
- Nambi V.E., Thangavel K., Shahir S., and Chandrasekar V., 2016b.** Colour kinetic during ripening of Indian mangoes. *Int. J. Food Properties*, 19, 2147-2155.
- Padda M.S., do Amarante C.V., Garcia R.M., Slaughter D.C., and Mitcham E.J., 2011.** Methods to analyze physico-chemical changes during mango ripening: A multivariate approach. *Postharvest Biol. Technol.*, 62, 267-274.
- Prasanna V., Prabha T.N., and Tharanathan R.N., 2007.** Fruit ripening phenomena – an overview. *Critical Reviews in Food Science and Nutrition*, 47, 1-19.
- Rungpichayapichet P., Mahayothee B., Nagle M., Khuwijitjaru P., and Müller J., 2016.** Robust NIRS models for non-destructive prediction of postharvest fruit ripeness and quality in mango. *Postharvest Biol. Technol.*, 111, 31-40.
- Saranwong S., Sornsrivichai J., and Kawano S., 2004.** Prediction of ripe-stage eating quality of mango fruit from its harvest quality measured nondestructively by near infrared spectroscopy. *Postharvest Biol. Technol.*, 31, 137-145.
- Saxena M. and Gandhi C.P., 2015.** *Indian Horticulture database 2014*. National Horticulture Board, Ministry of Agriculture, Government of India, New Delhi, India.
- Seymour G.B., Taylor J.E., and Tucker G.A., 1993.** *Biochemistry of fruit ripening*. Chapman and Hall, Malaysia.
- Stover R.H. and Simmonds N.W., 1987.** *Bananas*. Longman Scientific and Technical, Essex, England.
- Vásquez-Caicedo A.L., Sruamsiri P., Carle R., and Neidhart S., 2005.** Accumulation of all-trans- β -carotene and its 9-cis and 13-cis stereoisomers during postharvest ripening of nine Thai mango cultivars. *J. Agricultural Food Chemistry*, 53, 4827-4835.