Denitrification, organic matter and redox potential transformations in Cambisols

T. Włodarczyk¹*, Z. Stępniewska^{1,2}, and M. Brzezińska¹

¹Institute of Agrophysics, Polish Academy of Sciences, Doświadczalna 4, P.O. Box 201, 20-290 Lublin 27, Poland ²Catholic University of Lublin, Kraśnicka 102, 20-718 Lublin, Poland

Received June 7, 2003; accepted September 15, 2003

A b s t r a c t. Two greenhouse gases: N₂O and CO₂ as well as Corg and redox potential transformation were investigated. Fourteen Cambisols (0-10 cm) developed from sand, silt, loess, loam and clay characteristic for Poland were incubated under flood conditions in closed flasks at 20°C temperature for 7 days. According to denitrification activity the soils were divided into two groups - of lower (I) and higher (II) activity. Production of nitrous oxide lasting 3-5 days was followed by its absorption in the case of three soils out of fourteen. The total N2O amount reached from 3-91% of the initial nitrate - N content and was positively correlated with the Corg content. Diurnal N2O production was positively correlated with $C_{\rm org}$ and diurnal CO_2 emission. The N_2O absorption rate for the three soils was 0.16 mg $N_2 O\text{-}N\,kg^{-1}day^{-1}, 20.6$ mg N_2 O-N kg⁻¹day⁻¹ and 3.3 mg N_2 O-N kg⁻¹day⁻¹ in sandy, loamy and in the clay soil, respectively. The diurnal fluxes of carbon dioxide ranged from 1.24 to 9.96 mg of CO_2 -C kg⁻¹ d⁻¹ and from 26.8 to 144 mg of CO_2 -C kg⁻¹ for I and II group, respectively. Diurnal carbon dioxide production showed a high positive correlation with $C_{\mbox{\scriptsize org}}.$ The soils investigated were characterised by a very wide range of redox potential measured for the maximal cumulative N_2O emission from +417 to +233 mV. The beginning of N₂O emission from the light textured soils was observed above 400 mV while from the heavier textured soil below 400 mV. N2O emission was correlated with soil redox potential.

K e y w o r d s: N_2O emission, N_2O absorption, redox potential, soil respiration, CO_2 : N_2O ratio

INTRODUCTION

Soils are important sources of several greenhouse gases such as water vapour, CO₂, CH₄ and N₂O (Maljanen *et al.*, 2003; Xu-Ri *et al.*, 2003). In general, denitrification and chemolitotrophic nitrification are accepted as the dominant sources of N₂O in most terrestrial ecosystems (Firestone and Davidson, 1989; Martikainen *et al.*, 1993). In the field, both processes are often limited by substrate-availability (nitrate, ammonium), especially in N-limited forests (Gundersen and Rasmussen, 1990). The three main factors controlling soil denitrification rates are generally considered to be the concentration of oxygen, nitrate concentration and the availability of easily metabolizable organic matter (Tiedje, 1988; Horn *et al.*, 1994). These factors interact in a complicated manner with microorganisms on a microscale level in the soil, creating the large spatial and temporal variability in denitrification that has been observed in several studies (Parkin *et al.*, 1987; Svensson *et al.*, 1991).

Redox potential (Eh) is the aeration parameter characterising the intensity of soil redox transformations. The work of Smith *et al.* (1983) and Włodarczyk (2000) indicates that N₂O emission and absorption can occur in soil if the Eh falls bellow 300 mV.

In the present investigation two greenhouse gases, nitrous oxide and carbon dioxide emission and N_2O absorption as well as redox potential and the turnover of organic matter, were investigated under flooded soils amended with nitrate in order to better recognise the expected interrelations between these factors.

MATERIALS AND METHODS

Fourteen arable Polish top soils (0–10 cm) used in the study were Cambisols developed from different parent material: sand (Nos 39, 342, 434, 543, and 772), silt (Nos113, 224 and 984), loess (No. 672), loam (Nos 110, 328, 351 and 922) and clay (No. 947). Samples of these soils for the laboratory experiment were taken from the Bank of Soils situated in the Institute of Agrophysics in Lublin containing 1000 soil profiles. The 100 g portions of air-dry, sieved (1 mm sieve) soils were saturated with water and put on

^{*}Corresponding author's e-mail: teresa@demeter.ipan.lublin.pl

^{© 2003} Institute of Agrophysics, Polish Academy of Sciences

ceramic tension plates at 100 hPa (pF 2.0) for conditioning at 20°C in the dark. After 48 h of pre-incubation, moist soil samples (equal to 5 g of air-dry weight) were placed in 38 cm³ glass vessels and enriched with KNO₃ (2 ml of solution containing 1g NO₃⁻-N kg⁻¹) and distilled water (4 ml). The nitrate introduced equalled 200 mg NO₃⁻-N kg⁻¹ and the soil/water ratio was about 1:1 (w/w). The vessels with suspensions were tightly sealed with rubber stoppers and incubated in N₂ atmosphere with 2% (v/v) C₂H₂. The average concentration of O₂ in the headspace at the start of the incubation was 1.2%. Paraffin film was affixed to the stoppers to ensure a hermetic seal.

Soils were incubated at 20°C for seven days. After 1, 2, 3, 5, and 7 days the concentration of N_2O and CO_2 in the headspace was determined chromatographically using a Shimadzu GC-14 (Japan) fitted with a thermal conductivity detector at 60°C. Gas samples were separated on a 2 m column packed with a Porapak Q and a Molecular sieve and maintained at 40°C. Helium was the carrier gas flowing at a rate of 40 ml min⁻¹. The concentrations of N_2O and CO_2 were corrected for gas dissolved in the water by using the published values of the Bunsen absorption coefficient. Incubation vessels were prepared in three replicates. Additionally, a set of 12 samples obtained by the same procedure

T a ble 1. Basic properties of the investigated Cambisols

was prepared for each soil. These vessels were opened successively in three replications after 1, 2, 3, and 7 days of incubation to measure pH and redox potential (Eh), (Gliński and Stępniewski, 1995).

The diurnal N_2O and CO_2 production rates were calculated for the ranges of a linear increase of the headspace N_2O content only (as the production divided by the time).

The linear (y=a+bx), multiplicative (y=ax^b), exponential (y= e^{a+bx}) and logarithmic (y=alnx+b) models were used in the regression analysis and in each case the model with the highest R^2 was selected as the best fit for the experimental data.

RESULTS AND DISCUSSION

N₂O emission and absorption

The soils investigated showed a high variation in soil texture, organic carbon content, pH and endogenous nitrate content (Table 1).

 N_2O production for the soils amended with 200 mg $NO_3^{-}-N$ kg⁻¹ and in an anaerobic manner, incubated for 7 days, followed a specific and a different pattern. According to denitrification activity, the soils investigated were divided into two groups. Soils, which showed lower activity

Parent	Soil	Soil number	Soil fractions (%)					Native NO ₃ ⁻ -N	Native+added NO ₃ ⁻ -N
material	group		1-0.1	0.1-0.02 (mm)	< 0.02	рН	C_{org} (%)	(mg kg ⁻¹)	
Sand	Ι	342	70	11	19	7.07	0.44	29.90	229.9
	Ι	434	71	18	11	4.45	0.74	18.50	218.5
	Ι	772	76	11	13	4.50	0.49	13.70	213.7
	Ι	39	89	6	5	4.64	0.67	4.53	204.5
	II	543	57	23	20	5.99	0.88	9.70	209.8
Silt	Ι	224	81	8	11	6.32	0.32	6.47	206.5
	Ι	113	66	26	8	4.50	0.92	58.40	258.4
	II	984	16	48	36	4.87	1.24	7.87	207.9
Loam	Ι	351	71	8	21	7.53	0.57	17.90	217.9
	Ι	110	42	25	33	4.38	0.67	10.20	210.2
	II	328	48	22	30	6.03	0.77	9.46	209.5
	II	922	16	57	27	4.45	1.89	5.37	205.4
Clay	II	947	4	64	32	5.29	2.31	32.90	232.9
Loess	II	672	1	60	39	4.85	0.94	67.40	267.4

(I) where N₂O production was below 7 mg of N₂O-N kg⁻¹ d⁻¹ of soil, and higher activity (II), where N₂O production was up to 47.2 mg of N₂O-N kg⁻¹ d⁻¹ (Table 2). Eight soils belong to the first group; four of them are developed from sand (Nos 39, 342, 434 and 772), two soils are developed from loam (Nos 110 and 351) and two are developed from silt (Nos 113 and 224). Six soils belong to the second group; one is developed from silt (No. 984), two are developed from loam (Nos 328 and 922), one is developed from sand (No. 543), one is developed from loess (No. 672) and one is developed from clay (No. 947).

to group I and ranging from 208 to 270 mg of $NO_3^{-}-N kg^{-1}$ (Table 1). It seems probable that the next five soils (Nos 543, 984, 672, 328, and 922) had a low affinity to nitrate (Fig. 1b).

Soil No. 947 showed different denitrification activity. The N₂O production in the nitrate treatment followed first-order kinetics (Table 1) above 233 mg of initial NO₃⁻-N kg⁻¹ concentration (Fig. 1b). It seems that this soil had a higher affinity to nitrate than the rest of the soils investigated. In laboratory studies of potential denitrification, the NO₃⁻ concentration associated with this transition is thought to be relatively high, with values ranging from 40

Functional classification of soils	Diurnal N ₂ O emission (mg N ₂ O-N kg ⁻¹ d ⁻¹)	Eh (mV) at maximal denitrification	Diurnal CO ₂ emission (mg CO ₂ -C kg ⁻¹ d ⁻¹)	C _{org} (%)
Lower activity (I)	1.10-6.84	233-420	1.24–9.96	0.32-0.74
Higher activity (II)	9.40-47.2	237–391	3.83-32.6	0.77–2.31

The emission of nitrous oxide from the first group (I) of soils was very low and showed a small increase independently of any initial nitrate concentration (native + nitrate addition) during the seven days of incubation (Fig. 1a). The total cumulative emission of N2O ranged from 6.34 to 26.6 N_2 O-N mg kg⁻¹ and the diurnal fluxes of nitrous oxide from this group ranged from 1.10 to 6.84 mg of N_2 O-N kg⁻¹ d⁻¹ (Table 2). The conclusion was drawn that in the first group of soils, denitrification was independent of the NO₃⁻ concentration with zero order reaction ranging between about 205 to 230 mg of initial $NO_3^{-}-N \text{ kg}^{-1}$ concentration (Table 1). It has been widely reported that with the increasing concentration of soil NO_3^- , denitrification changes from being dependent on NO_3^- , with the first order of Michaelis-Menten kinetics, to being independent of NO_3^{-} , that is, following zero order kinetics (Scholefield et al., 1997). The low response of the soils investigated on the relatively high nitrate addition suggested that most of them had a low affinity (defined as the reciprocal of the Michaelis constant -K_M) to nitrate which is the reaction of the medium for a given substrate.

The similar course of the cumulative curve of N_2O emission showed the second group (II) of soils as having a higher activity, except for one (No. 947); however, this group was characterised by a higher nitrous oxide production. The total cumulative emission of N_2O ranged from 65.8 to 187.4 N_2O -N mg kg⁻¹ and the highest diurnal fluxes of nitrous oxide from this group was above 47 mg of N_2O -N kg⁻¹ d⁻¹ (Table 2). In the case of the second group of soils the denitrification changes were independent of NO_3^- concentration with zero order kinetics in a concentration similar



Fig. 1. The course of cumulative nitrous oxide content (mean values + standard deviations) in the headspace during the incubation of the two soil groups: 8 soils with lower activity (a) and 6 soils with higher activity (b). A discontinuous line denotes soil where N_2O absorption was observed.

(Knowles, 1981) to 100 mg of N kg⁻¹ soil (Aulakh *et al.*, 1992) being quoted as typical. Scholefield *et al.*, (1997) found that denitrification in Stagno-Dystric Gleysol responds to increasing NO₃⁻ supply within the range 0–150 kg of N ha⁻¹ (equivalent approximately to 0–150 mg kg⁻¹). In our investigations, the threshold of NO₃⁻ concentration is higher than 200 mg kg⁻¹ (600 kg of N ha⁻¹).

The total cumulative N_2O emission expressed as a percentage of the initial nitrate content was distinctly varied in the soil under investigation. As far as the percentage of the initial nitrate – N converted to nitrous oxide is concerned, the data presented in Fig. 2 indicates that it varied from 2.8 to about 91%. The differences between the particular soil groups was clear and ranged from 2.8 to 10.4% and from 24.6 to 91.2% for the first and second group, respectively.

After a day of the maximum content of N_2O , its stepwise disappearance from the headspace was observed. The absorption rates of N_2O were distinctly different in the groups of soil. As can be seen from Fig. 2, only three soils showed the ability to absorb N_2O , one from group I (No. 342) and two from group II (Nos 922 and 947). The nitrous oxide generated underwent absorption during the incubation time in a different manner. The sandy soil (No. 342) absorbed about 10% of the N₂O emitted. The loamy soil (No. 922) showed the highest N₂O absorption (about 22%) while the lowest, (about 7%) was observed in the case of the clay soil (No. 947). The soils under investigation showed a distinct differentiation in the diurnal nitrous oxide absorption (0.16 for sandy soil, 3.3 for clay, and 20.6 mg N₂O-N kg⁻¹ d⁻¹ for the loamy soil).

Organic matter turnover

The lower cumulative emission of CO_2 on seventh day of incubation was observed in the first group (I) of those soils studied, from 8.7 to 50.4 mg of CO_2 -C kg⁻¹ (Fig. 3a) compared with the second group. The diurnal fluxes of carbon dioxide (calculated only for the ranges of a linear increase of the headspace N₂O content) observed in this group ranged from 1.24 to 9.96 mg of CO₂-C kg⁻¹ d⁻¹ (Table 2).





Fig. 2. Percentage of N-forms on the last day of incubation $(NO_3^--N, N_2O-N \text{ emitted and } N_2O-N \text{ absorbed})$. 8 soils with lower activity (a) and 6 soil with higher activity (b).

Fig. 3. The course of cumulative carbon dioxide content (mean values + standard deviations) in the headspace during the incubation of the two soil groups: 8 soils with lower activity (a) and 6 soils with higher activity (b). A discontinuous line denotes soil where N_2O absorption was observed.

The low CO_2 production was probably connected with the low organic matter content (0.32–0.74%).

The soils of the second group (II) (Fig. 3b), showed approximately a four times higher CO_2 production which corresponded to the higher C_{org} content (0.77–2.31%). Carbon dioxide emission on the seventh day of incubation ranged from 26.8 to 144 mg of CO_2 -C kg⁻¹. The diurnal flux of carbon dioxide from this group ranged from 3.83 to 32.6 mg of CO_2 -C kg⁻¹ d⁻¹ (Table 2).

 CO_2 production from most of the soils investigated during the first three days of anaerobic incubation followed linearly and independently of the denitrification activity of the soils studied (Fig. 3a,b). C mineralization in these soils apparently followed zero kinetics. After the third day of incubation, CO_2 production slightly decreased in one soil with higher C_{org} content (No. 922). Any prolonged incubation (above three days) would have reduced the easily available organic C substrate. The adaptation of the microorganisms over two days to more heavily decomposable organic matter resulted in an increase of CO_2 emission (Fig. 3b).

The results clearly indicate the influence of soil C_{org} on CO_2 emission. Diurnal CO_2 emission (Fig. 4) was positively correlated with the C_{org} content ($R^2=0.91***$).

The very small CO₂ emission was accompanied by a very small N₂O emission in the case of the first group of soils while a proportionally higher N₂O and CO₂ emission was observed in the second group of soils. Diurnal carbon dioxide production was closely related to diurnal N₂O emission (Fig. 5) and showed a high positive correlation (R^2 =0.952***). Other authors (Dendooven and Anderson, 1995; Dendooven *et al.*, 1996; Swerts *et al.*, 1996) obtained similar results. They found that total denitrification rates.

It should be emphasized that the diurnal nitrous oxide formed due to denitrification (Fig. 6) and the percentage of nitrate denitrified to nitrous oxide (Fig. 7) were positively related to the C_{org} content of the soil ($R^2=0.95^{***}$ and $R^2=$ 0.60^{***} , respectively). The results proved the close connection of N and C transformation pathways in the metabolism of microorganisms inhabiting the soil system.

The soils tested varied in molecular $CO_2:N_2O$ ratio (Table 3). As a rule, the molecular ratio between CO_2 and the N_2O production systematically decreased in the soils included in the first group (I). The widest ratios (2.7–14.5) were observed during the first day of incubation and probably mainly as a result of an excess of CO_2 production coinciding with a lag in N_2O production at the beginning of incubation. Swertes *et al.* (1996) found that denitrification rates during the first phase of denitrification are determined by the pre-existing enzymatic capacity to denitrify rather than by the availability of nitrate or carbon. The wide $CO_2:N_2O$ ratio (more than 2) at the beginning of our incubation might suggest that denitrification and fermentative CO_2 production took place simultaneously. Generally, the rate of



Fig. 4. Diurnal CO₂-C emission versus of C_{org} content under consideration.



Fig. 5. Diurnal N₂O-N emission versus diurnal CO₂-C emission under consideration.



Fig. 6. Diurnal N_2 O-N emission versus of C_{org} content under consideration.



Fig. 7. Total headspace N_2O -N expressed as percent of the initial NO_3^- -N content versus C_{org} content.

 N_2O and CO_2 emission was apparently higher in more heavily textured soils (developed from clay and loam). Notwithstanding, those soils where sand is dominant as the parent material showed a low capacity to denitrify. It seems that the wide molecular ratio was connected with the very low denitrification activity observed in the soils investigated and with the conversion of polymerised carbon compounds into readily available forms during drying, especially sand soils. Burford and Bremner (1975) found that the capacity to denitrify was directly related not just to the total, but also to the carbon which was water-soluble and able to be mineralized in reaction mixtures of sand, silt, and clay incubated at 20°C for 7 days. During subsequent incubation, the $CO_2:N_2O$ ratios were lower on the seventh day of incubation reaching values from 3.6 to 0.4, indicating that most of the soil investigated had adapted to the anaerobic conditions and that the denitrification process had pre-dominated. The simultaneous exhaustion of readily available C stopped CO₂ production (Fig. 3). Pidello *et al.* (1996) studied N₂O and CO₂ emission under different redox potential conditions. The ratio between CO₂ and N₂O production was continuously more than 2 suggesting that denitrification and fermentative CO₂ production had taken place simultaneously.

The molecular ratio between the CO_2 and the N_2O production, which well reflected the capacity for denitrification, showed a completely different course in the second group (II) of the soils investigated (with the exception of soil No. 984). The ratios were less diversified at the beginning of incubation (i.e., 3.2–0.3) and after the second day of incubation decreased below 2 (Table 3) indicating that N_2O was the dominant product of denitrification. According to stoichiometric calculations, the relationship between N_2O production and CO_2 equals 2 when N_2O is the only product of denitrification (Delwiche, 1981). Soils classified in group II were characterised by a low molecular ratio between the CO_2 and the N_2O production and the probably higher level of pre-existing enzymes which participated in dissimilar nitrate reduction.

It is concluded from our study that most of the soils were not well adapted to anaerobic conditions and showed a low capacity to denitrify; this is probably connected partly with the soil texture. The rest of the soils studied had a high capacity for denitrification. The molecular ratio between CO_2 and N_2O production amply reflected the differences in the soil's capacity to denitrify where nitrate is the only electron acceptor.

Table 3. The molecular ratio between the CO₂ and the N₂O production (CO₂:N₂O) calculated for the investigated soils

Functional classification of soils	Soil No.	Day of incubation				
		1	2	3	5	7
	342	6.5	2.7	1.7	1.7	1.7
	434	8.8	4.5	3.7	3.4	3.6
	772	14.5	3.4	3.1	4.5	3.2
Lower activity (I)	39	2.7	1.6	2.0	1.7	1.7
Lower activity (1)	113	5.8	2.9	1.5	2.1	1.9
	224	2.7	1.2	0.4	0.6	0.4
	351	7.5	3.2	1.9	1.7	1.4
	110	12.1	3.8	2.0	2.0	1.7
	543	1.3	1.3	0.9	1.0	0.8
	984	3.2	2.3	1.2	1.4	1.0
Higher activity (II)	947	0.9	0.7	0.9	1.5	1.5
Higher activity (II)	672	1.2	1.2	1.0	1.2	1.1
	328	0.3	0.3	0.2	0.3	0.4
	922	1.4	1.1	1.2	0.7	0.7

Nitrous oxide content versus soil redox potential

Several factors are known to influence the rate of denitrification directly or indirectly. One of the most important is redox potential (Eh). Under anaerobic conditions, the redox potential systematically dropped during incubation of the flooded soils (except for soil No. 672). The redox potential at the beginning of the incubation ranged from +301 to +434 mV and from +342 to +400mV for the first and the second group of soils, respectively (Table 4). It should be emphasised that the Eh value above 400 mV corresponded with the lowest N2O emission after first day of incubation. It seems probable that it is the threshold value for the beginning of the denitrification process under the conditions investigated. These soils are characterised by their high resistance to reduction processes. Other laboratory studies have shown that the high redox potential of the soils investigated at low water tensions is indicative of their high redox buffering capacity (t₃₀₀) Stępniewski et al. (2000). Gliński and Stepniewska (1986) defined index t_{300} as the time needed to lower the soil redox potential under flood conditions at a fixed temperature to a level of 300 mV.

After seven days of incubation, the Eh values apparently decreased in the soils investigated (by 3-121 mV) with the exception of one where the Eh value increased insignificantly from +377 to +391 mV (soil No. 673).

According to the data in Table 2 the highest cumulative N_2O emission from the first group of soil corresponded to the Eh value from +233 to +417 mV. A slightly lower redox potential value was noticed in the case of soils with higher

denitrification activity and ranged from 237 to 391 mV. The amount of denitrified N₂O was the highest in the range of Eh values from about +250 to +319 mV. Generally, the soils investigated were characterised by a very wide range of redox potential measured for maximal cumulative N₂O emissions (+417 to +233 mV) (Table 2).

The value of redox potential corresponding to the absorption of N₂O was below +340 mV (Table 3). Patrick (1960) showed that denitrification can occur when the redox potential of soil has decreased to below about +340 mV. Masscheleyn *et al.* (1993) reported N₂O emission from rice paddy soils with various redox potentials, ranging from +500 to -250 mV. Two maximums for N₂O evolution were found, at +400 mV where nitrification was the source and at 0 mV where N₂O was produced by denitrification.

The amount of nitrous oxide formed due to denitrification showed a negative correlation with the Eh value in most of the soils investigated (Table 5). This close relationship between denitrification and redox potential was confirmed by the correlation between the molecular $CO_2:N_2O$ ratio and Eh (Table 6). Pidello *et al.* (1996) studied N_2O and CO_2 emission under conditions of different redox potentials. The ratio between the CO_2 and the N_2O production was correlated with the Eh value.

Our results incline us to the conclusion that the threshold value of the redox potential for the emission of N_2O is very wide for different soils and depends on many factors. The beginning of N_2O emission from the light textured soils developed from sand was observed above +400 mV while this was below +400 mV for the heavily textured soils.

Functional classification of soils	Soil No. –	Day of incubation			
		1	2	3	7
	342	379	364	339	315
	434	434	429	430	412
	772	413	410	229	311
Lower activity (I)	39	410	385	235	299
Lower activity (1)	113	420	400	385	417
	224	301	311	212	233
	351	326	295	234	266
	110	392	373	313	308
	543	356	377	232	237
	984	399	390	397	319
Ui al an a stinita (U)	947	342	331	280	221
Higher activity (II)	672	377	378	389	391
	328	356	350	352	334
	922	400	393	333	302

T a b l e 4. The redox potential (mV) of the investigated soils

Soil No.	Function	R^2	n
	I group		
342	y = -6.1335x + 382.17	0.779***	9
434	$y = 436.41e^{-0.0071x}$	0.299*	15
772	Not significant		
39	$y = -64.12 \ln(x) + 457.72$	0.394**	15
113	Not significant		
224	$y = -31.872 \ln(x) + 316.51$	0.622***	15
351	$y = -22.707 \ln(x) + 297.17$	0.512**	15
110	$y = -29.26 \ln(x) + 394.33$	0.809***	15
	II group		
543	$y = 912.64x^{-0.3324}$	0.639***	15
984	$y = 405.88e^{-0.0021x}$	0.810***	15
947	y = 0.9675x + 381.07	0.496**	12
672	Not significant		
328	y = -0.3429x + 360.05	0.411**	15
922	$y = -31.756 \ln(x) + 487.33$	0.651***	12

T a ble 5. Values for the correlation coefficient (R^2) obtained between cumulative content of N_2O -N in the headspace of gas for the phase of emission and redox potential

Significance level: ***p<0.001, **p<0.01, *p<0.05.

T a b l e 6. Values for the correlation coefficient (R^2) obtained between the molecular ratio of CO₂:N₂O for the diurnal N₂O emission and redox potential

Group of soil	Function	R ²	n	
I group	$y = 8E - 06x^{2.1977}$	0.356***	108	
II group	y = 0.0045x - 0.484	0.107**	78	

CONCLUSIONS

1. The diurnal flux of nitrous oxide was no more than 6.84 mg of N₂O-N kg⁻¹ d⁻¹ in the first group of soils (I) with lower denitrification activity and was above 47 mg of N₂O-N kg⁻¹ d⁻¹ in the second group of soils (II) with higher activity.

2. The diurnal fluxes of carbon dioxide ranged from 9.96 to 32.6 mg of CO_2 -C kg⁻¹ d⁻¹ in the first (I) and second (II) groups of soils, respectively.

3. The total cumulative N_2O emission expressed as a percentage of the initial nitrate content varied from 2.8 to about 91%.

4. Three soils showed an ability to absorb N_2O and absorbed from 7 to 22% of the nitrous oxide emitted.

5. Diurnal $\rm N_2O$ and $\rm CO_2$ production was positively correlated with $\rm C_{org}.$

6. The beginning of N_2O emission from the light textured soils was observed above +400 mV while from the heavier textured soil this was below +400 mV.

 $7. N_2O$ emission was negatively correlated with the soil redox potential.

REFERENCES

- Aulakh M.S., Doran J.W., and Mosier A.R., 1992. Soil denitrification-significance, measurement, and effects of management. In: Advances in Soil Science (Ed. Stewart B.A.). Springer, New York, 1–75.
- **Burford J.R. and Bremner J.M., 1975.** Relationships between the denitrification capacities of soils and total, water soluble and readily decomposable soil organic matter. Soil Biol. Biochem., 7, 389–394.
- **Delwiche C.C., 1981.** The nitrogen cycle and nitrous oxide. In: Denitrification, nitrification and atmospheric nitrous oxide (Ed. C.C. Delwiche). John Wiley, New York, 1–15.
- Dendooven L. and Anderson J.M., 1995. Maintenance of denitrification potential in pasture soil following anaerobic events. Soil Biol. Biochem., 10, 1251–1260.
- **Dendooven L., Pemberton E., and Anderson J.M., 1996.** Denitrification potential and reduction enzyme dynamics in a Norway spruce plantation. Soil Biol. Biochem., 28, 151– 157.
- **Firestone M.K. and Davidson E.A., 1989.** Microbiological basis of NO and N₂O production and consumption in soil. In: Exchange of the trace gases between terrestrial ecosystems and the atmosphere (Eds M.O. Andrea, D.S. Schimel). Report for the Dahlem Workshop on Exchange of Gases

between terrestrial Ecosystems and the Atmosphere, Berlin, J. Wiley and Sons, 7–22.

- Gliński J. and Stępniewska Z., 1986. An evolution of soil resistance to reduction processes. Pol. J. Soil Sci., 19, 15–19.
- Gliński J. and Stępniewski W., 1985. Soil Aeration and Its Role for Plants. CRC Press Inc., Boca Raton, Florida.
- **Gundersen P. and Rasmussen L., 1990.** Nitrification in forest soils: effects from nitrogen deposition on soil acidification and aluminium release. Reviews of Environmental Contamination and Toxicology, 113, 1–45.
- Horn R., Stępniewski W., Włodarczyk T., Walenzik G., and Eckhardt F.E.W., 1994. Denitrification rate and microbial distribution within homogenous model soil aggregates. Int. Agrophysics, 8, 65–74.
- Knowles R., 1981. Denitrification. In: Terrestrial Nitrogen Cycles, (Eds F.E. Clark, T.D. Rosswall). Ecological Bulletin, Stockholm, 33, 315–329.
- Maljanen M., Liikanen A., Silvola J., and Martikainen P.J., 2003. Nitrous oxide emission from boreal organic soil under different land-use. Soil Biol. Biochem., 35, 1–12.
- Martikainen P.J., Lehtonen M., Lang K., De Boer W., and Ferm A., 1993. Nitrification and nitrous oxide production potentials in aerobic soil samples from the soil profile of a Finnish coniferous site receiving high ammonium deposition. FEMS Microbiology Ecology, 13, 113–122.
- Masscheleyn P.H., DeLaune R.D., and Patrick W.H., 1993. Methane and nitrous oxide emissions from laboratory measurements of rice soil suspension – effect of soil oxidation-reduction status. Chemosphere, 26, 251–260.
- Parkin T.B., Starr J.L., and Meisinger J.J., 1987. Influence of sample size on measurement of soil denitrification. Soil Sci. Soc. Am. J., 51, 1492–1501.
- Patrick W.H., 1960. Nitrate reduction rates in a submerged soil as affected by redox potential. Trans. 7th Int. Congr. Soil Sci., 2, 494.

- Pidello A., Menéndez L., and Perotti E.B.R., 1996. Saccharidic compounds as soil redox effectors and their influence on potential N₂O production. Biol. Fertil. Soils, 23, 173–176.
- Scholefield D., Hawkins J.M.B., and Jackson S.M., 1997. Use of a flowing helium atmosphere incubation technique to measure the effects of denitrification controls applied to intact cores of a clay soil. Soil Biol. Biochem., 9/10, 1337–1344.
- Smith C.J., Wright M.F., and Patrick W.H. jr., 1983. The effect of soil redox potential and pH on the reduction and production of nitrous oxide. J. Environm. Qual., 12, 186–188.
- Stępniewski W., Stępniewska Z., Gliński J., Brzezińska M., Włodarczyk T., Przywara G., Varallyay G., and Rajkaj K., 2000. Dehydrogenase activity of some Hungarian soils as related to their water and aeration status. Int. Agrophysics, 14, 365–376.
- Svensson B.H., Klemedtson L., Simkins S., Paustian K., and Rosswall T., 1991. Soil denitrification in three cropping systems characterised by differences in nitrogen and carbon supply. I. Rate distribution frequencies, comparison between systems and seasonal N-losses. Plant and Soil, 138, 257–271.
- Swerts M., Merckx R., and Vlassak K., 1996. Influence of carbon availability on the production of NO, N₂O, N₂ and CO₂ by soil cores during anaerobic incubation. Plant and Soil, 181, 145–151.
- **Tiedje J.M., 1988.** Ecology of denitrification and dissimilatory nitrate reduction to ammonium. In: Biology of anaerobic microorganisms (Ed. A.J.B. Zehner). John Wiley, New York, 179–244.
- Włodarczyk T., 2000. N₂O emission and absorption against a background of CO₂ in Eutric Cambisol under different oxidation-reduction conditions (in Polish). Acta Agrophysica, 28.
- Xu-Ri, Wang M., and Wang Y., 2003. Using a modified DNDC model to estimate N₂O fluxes from semi-arid grassland in China. Soil Biol. Biochem., 35, 615–620.